



Original scientific paper

ZnO/1-hexyl-3-methylimidazolium chloride paste electrode, highly sensitive lorazepam sensor

Seddigheh Chenarani¹, Mahmoud Ebrahimi^{1,✉}, Vahid Arabali² and Safar Ali Beyramabadi²

¹Department of Chemistry, Mashhad Branch, Islamic Azad University, Mashhad, Iran

²Department of Chemistry, Sari Branch, Islamic Azad University, Sari, Iran

Corresponding author: ✉ m.ebrahimi@mshdiau.ac.ir; Tel.: +98-9155095175h; Fax: +98-51-3663-0781

Received: October 30, 2022; Accepted: December 5, 2022; Published: January 18, 2023

Abstract

The measurement of pharmaceutical compounds in biological fluids is considered an effective way to evaluate their effectiveness. On the other hand, lorazepam is a drug with good efficiency in treatment and some side effects, which measurement is very important. In this study, the ZnO nanoparticle was synthesized as an electrocatalyst by chemical precipitation method. Then, a simple modification on paste electrode (PE) by ZnO nanoparticle (ZnO-NPs) and 1-hexyl-3-methylimidazolium chloride (HMIImCl) was made and a new sensor was used for sensing of lorazepam. The HMIImCl/ZnO-NPs/PE showed catalytic behavior on oxidation signal of lorazepam and improved its signal about 2.17 times compared to unmodified PE. On the other hand, oxidation potential of lorazepam was reduced about 110 mV at surface of HMIImCl/ZnO-NPs/PE compared to unmodified PE that confirm accelerating the electron exchange process after modification of sensor by HMIImCl and ZnO-NPs as powerful catalysts. The HMIImCl/ZnO-NPs/PE was used for monitoring of lorazepam in water and injection samples and results showed recovery data 98.5 to 103.5 % that are acceptable for a new sensor.

Keywords

Pharmaceutical sensor; modified sensor; nano-catalyst; ionic liquid

Introduction

Medicines, as one of the most important substances used in human life, have many positive and negative effects, which have attracted the attention of many researchers [1,2]. Although drugs play an important role in the treatment of diseases, the harmful effects of some of them on the body as well as the harmful effects on the environment have caused many studies to be done on this group of compounds [3]. Measuring medicinal compounds is one of the most important studies in this field and can provide a lot of information during patient treatment or its role in environmental pollution [4,5]. In between of analytical methods in sensing of pharmaceutical compounds [6-10], electrochemical

methods showed more attentions due to many advantages such as easy modification for sensitive sensing, low cost and wide range applications [11].

The lorazepam is one of famous medicines used to treat anxiety [12]. There are several reports of adverse effects of lorazepam use such as dizziness, tiredness and weakness [13]. Therefore, monitoring of this medicine is very important during treatment [14]. Due to high over-voltage and low redox signal of lorazepam, the sensing of it is so hard with usual electrochemical sensors [11]. To overcome to this problem, modification and amplification of electrochemical sensors is necessary [15]. Modification of electrodes is one of the proven solutions to increase the sensitivity and selectivity of electrochemical sensors [16-21]. Different types of mediators such as polymers, nanomaterials, ionic liquids, MOF *etc.* were suggested for modification and amplification of electrochemical sensors in recent years [22-30].

Nanotechnologies opened a new approach in science and improve many of chemical and physical properties of materials [31-36]. With this way, nanomaterials were selected as first choice in different branches of science [37-42]. Due to high electrical conductivity of some nanomaterials such as metal nanoparticles and carbon-based nanomaterials, they were used for modification of electrochemical sensors [43-45]. On the other hand, ionic liquids showed good advantage as binder for fabrication of paste electrode and improved electrical conductivity of modified paste electrodes [46-51].

In this research work, the HMIImCl/ZnO-NPs/PE was fabricated and used as new approach for monitoring of lorazepam with good limit of detection compared to previous suggested electrochemical sensors. The HMIImCl/ZnO-NPs/PE showed acceptable analytical data in sensing of lorazepam in real samples with recovery data 98.5 - 103.5%. Modification of electrode improved oxidation current of lorazepam about 2.17 times and reduced its oxidation potential for about 110 mV compared to unmodified electrode.

Experimental

Materials and instruments

Zinc nitrate hexahydrate and sodium hydroxide were purchased from Merck Company and used for synthesis of ZnO nanoparticles. 1-Hexyl-3-methylimidazolium chloride, graphite powder, paraffin oil and diethyl ether were purchased from Sigma-Aldrich Company and used for fabrication of paste electrode. Phosphoric acid purchased from Merck was used for preparation phosphate buffer solution. The *I* - *V* signals were recorded by Vertex – Ivium (potentiostat/galvanostat) connected with HMIImCl/ /ZnO-NPs/PE as working electrode, Ag/AgCl as reference electrode and Pt wire as counter electrode.

Synthesis of ZnO nanoparticles

The 100 mL zinc nitrate hexahydrate (0.5 M) were stirred in Erlenmeyer flask for 15 min and then 100 mL sodium hydroxide (0.5 M) were added dropwise and stirred form 30 min. White precipitate of zinc hydroxide was washed for 10 times by distilled water and then dried ate 100 °C for 16 h. The white powder was calcinated at 250 °C for 4 h and ZnO nano-powder was obtained.

Fabrication of HMIImCl/ZnO-NPs/PE

For fabrication of HMIImCl/ZnO-NPs/PE; 90 mg ZnO-NPs + 910 mg graphite powder were mixed in mortar and pestle and 15 mL diethyl ether was superimposed. After evaporation of diethyl ether, the paraffin oil + HMIImCl with ratio of (7:3 vol.%) were used as binders and sample hand mixed for 1 h. The HMIImCl/ZnO-NPs paste was inserted into the end of a glass tube for fabrication of HMIImCl/ZnO-NPs/PE in the presence of copper wire.

Results and discussion

Electrochemical investigations

The electrochemical behavior of lorazepam was investigated in the pH range 5.0 to 7.0 and results are shown in Figure 1.

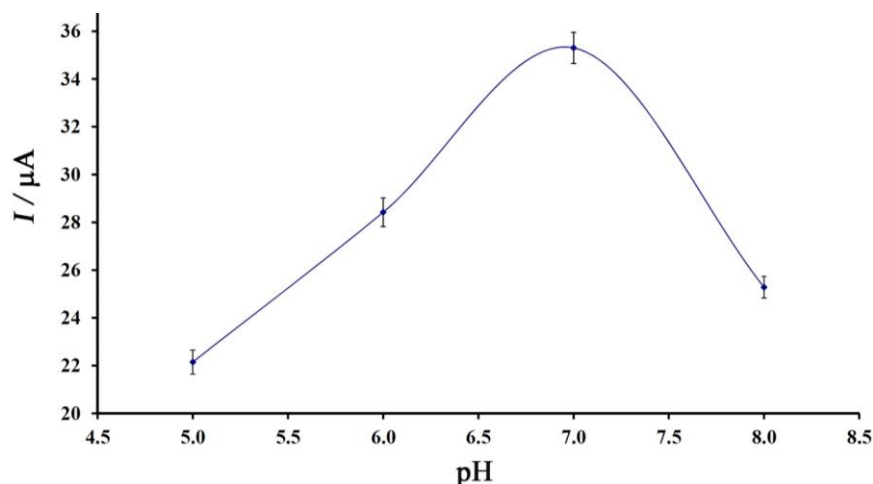


Figure 1. Oxidation peak current-pH diagram for sensing of 300 μM lorazepam using HMImCl/ZnO-NPs/PE as electroanalytical sensor ($n=4$)

As can be seen, with increasing pH from 5.0 to 7.0, the oxidation signal of 300 μM lorazepam increased and then decreased. Therefore, pH 7.0 was selected as optimum condition for monitoring of lorazepam using HMImCl/ZnO-NPs/PE as electroanalytical sensor.

The oxidation signal of 200 μM lorazepam was recorded at surface of PE (Figure 2a), ZnO-NPs/PE (Figure 2b), HMImCl/PE (Figure 2c) and HMImCl/ZnO-NPs/PE (Figure 2d), respectively.

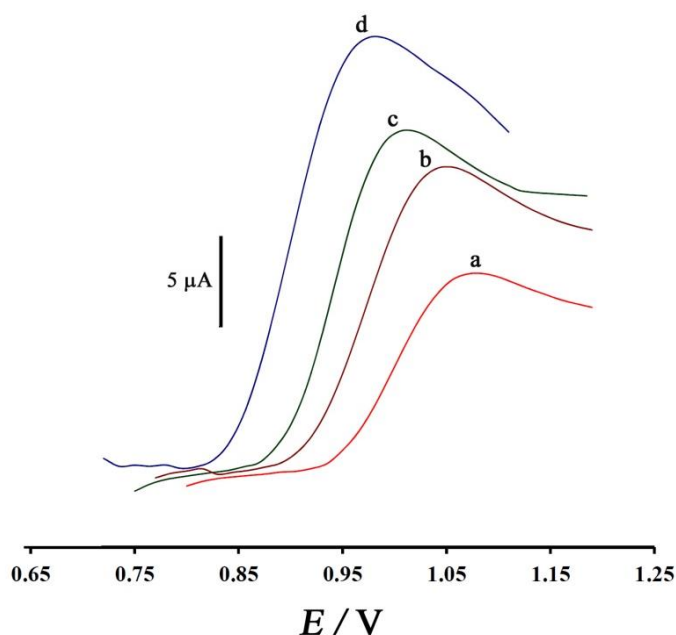


Figure 2. Linear sweep voltammograms of 200 μM lorazepam at surface of PE (a), ZnO-NPs/PE (b), HMImCl/PE (c) and HMImCl/ZnO-NPs/PE (d); pH 7.0

The oxidation peak currents of 10.85 μA , 16.53 μA , 18.55 μA and 23.6 μA were detected for monitoring of 200 μM lorazepam at surface of PE, ZnO-NPs/PE, HMImCl/PE and HMImCl/ZnO-NPs/PE, respectively. As can be seen, with moving PE to HMImCl/ZnO-NPs/PE, the oxidation current

of lorazepam increased due to high electrical conductivity and synergic effect of two conductive mediators. This result confirms synergic effect of two mediators after modification of PE and fabrication of a highly sensitive voltammetric sensor to monitoring of lorazepam.

The linear sweep voltammograms of 300 μM lorazepam at surface of HMIImCl/ZnO-NPs/PE and in the scan rate range 10 to 100 mV/s were recorded and signals are presented in the inset of Figure 3.

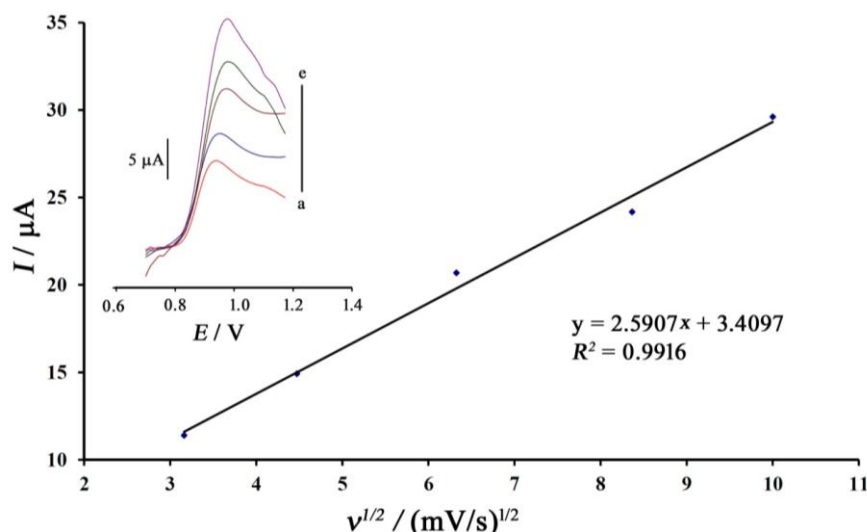


Figure 3. Peak current vs. $v^{1/2}$ plot for oxidation of 300 μM lorazepam at surface of HMIImCl/ZnO-NPs/PE. Linear sweep voltammograms of 300 μM lorazepam at scan rates; a) 10; b) 20; c) 40; d) 70 and e) 100 mV / s ($n=4$)

As can be seen, a linear relation between oxidation signals of 300 μM lorazepam and $v^{1/2}$ with equation $I = 2.5907 v^{1/2} + 3.4097$ ($R^2 = 0.9916$) that confirms diffusion process [52-55] to lorazepam oxidation at surface of HMIImCl/ZnO-NPs/PE. On the other hand, positive shift in oxidation potential of lorazepam with increase in scan rate confirms kinetic limitation in redox reaction of this drug.

The Tafel plot relative to oxidation of 300 μM lorazepam at scan rate 10 mV/s are shown in Figure 4. Using Tafel equation and Tafel slope, the value of α was calculated 0.33, that confirms irreversible behavior to redox reaction of lorazepam.

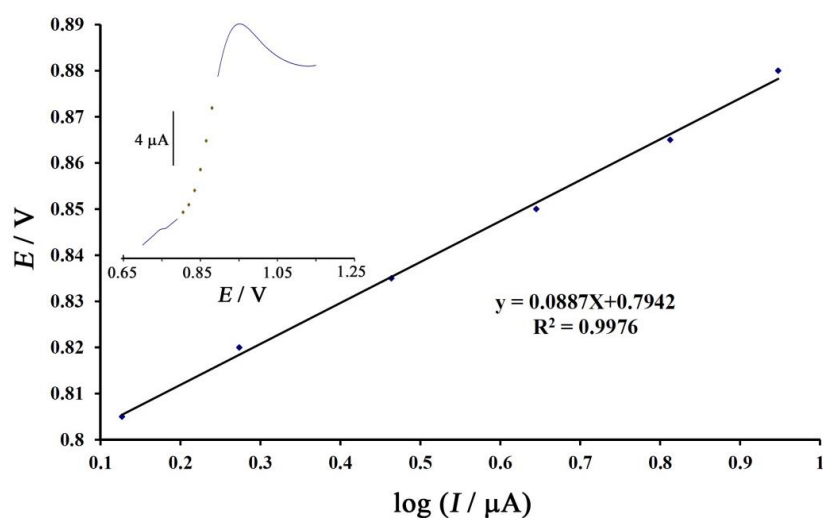


Figure 4. Tafel plot of 300 μM lorazepam at surface of HMIImCl/ZnO-NPs/PE with scan rate 10 mV/s

Analytical parameters

Linear dynamic range (LDR) and limit of detection (LOD) of proposed system for sensing of lorazepam was investigated by square wave voltammetric method (Figure 5 inset).

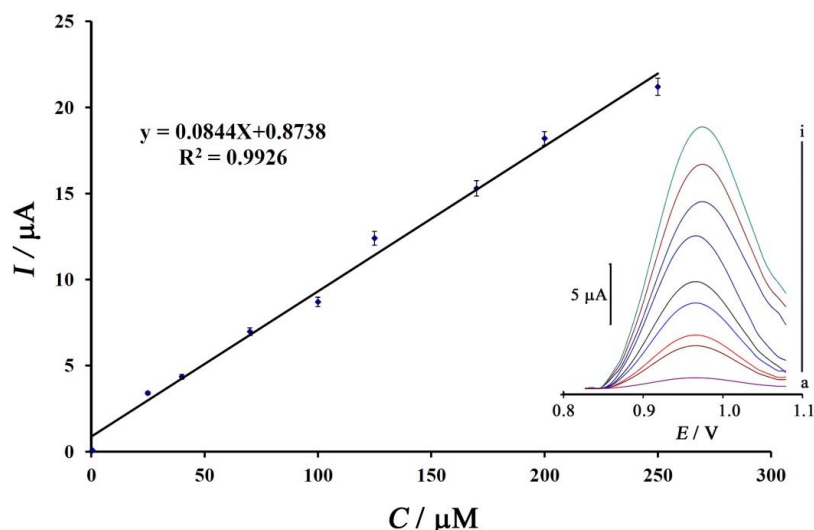


Figure 5. Current – concentration curve for monitoring of lorazepam using HMImCl/ZnO-NPs/PE. Inset) square wave voltammograms of lorazepam at surface of HMImCl/ZnO-NPs/PE at concentrations of 1) 0.5; 2) 25; 3) 40; 4) 70; 5) 100; 6) 125; 7) 170; 8) 200 and 9) 250 μM ($n=4$)

A linear relation in the concentration range 0.5 to 250 μM with equation $I = 0.0844C + 0.8738$ ($R^2 = 0.9926$) was detected for sensing of lorazepam using HMImCl/ZnO-NPs/PE as electro-analytical sensor. The detection limit 0.1 μM was reported for sensing of lorazepam using HMImCl/ZnO-NPs/PE in this study.

Stability, selectivity and real sample analysis

The stability of HMImCl/ZnO-NPs/PE for monitoring of 200 μM lorazepam was investigated in period time 70 days. Results shown in Figure 6 confirm that HMImCl/ZnO-NPs/PE has good stability for sensing of lorazepam in 2 months.

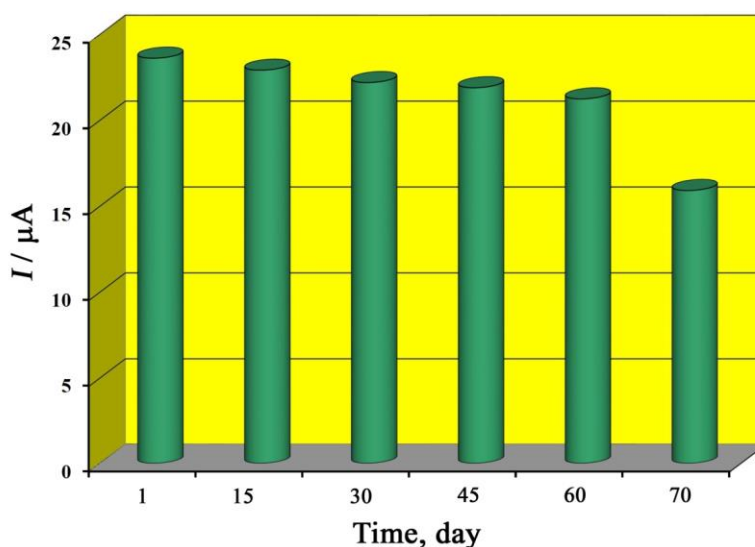


Figure 6. Current – days diagram for oxidation of 200 μM lorazepam at surface of HMImCl/ZnO-NPs/PE

On the other hand, selectivity of HMImCl/ZnO-NPs/PE in sensing of 15 μM lorazepam was investigated and results with acceptable error 5 % are reported in Table 1. As can be seen in Table 1, there is not any important interference observed for monitoring of 15 μM lorazepam using HMImCl/ZnO-NPs/PE and this sensor showed good selectivity in monitoring of lorazepam.

In the final step, ability of HMImCl/ZnO-NPs/PE in monitoring of lorazepam in water, dextrose saline and injection samples was checked, and results are reported in Table 2. As can be seen, the

HMIImCl/ZnO-NPs/PE detected lorazepam with recovery range 98.5 to 103.5 % that are acceptable values for a new sensor.

Table 1. Interference study results for monitoring 15 μM lorazepam using HMIImCl/ZnO-NPs/PE as sensor

Species	Tolerant limits ($W_{\text{substance}}/W_{\text{lorazepam}}$)
Cl^- , Br^- , Ca^{2+} , K^+ , Na^+	1000
Glucose	700
Starch	Saturation

Table 2. Application of HMIImCl/ZnO-NPs/PE for sensing of lorazepam in real samples

Sample	C / μM			Recovery, %
	Added	Expected	Founded	
Water	---	---	<LOD	---
	10.00	10.00	10.35 $\pm$ 0.54	103.5
Injection	---	2.00	2.03 $\pm$ 0.05	---
	10.00	12.00	11.82 $\pm$ 0.45	98.5
Dextrose saline	---	---	<LOD	---
	10.00	10.00	10.31 $\pm$ 0.51	103.1

Conclusions

In this study, a new and simple analytical plan was described for monitoring of lorazepam in aqueous solution. The suggested sensor (HMIImCl/ZnO-NPs/PE in this case) showed good selectivity for monitoring of lorazepam. The pH 7.0 was selected as optimum condition in voltammetric analysis. In addition, HMIImCl/ZnO-NPs/PE was successfully used to monitor of lorazepam in the concentration range 0.5 to 250 μM with detection limit 0.1 μM . The modification of PE by HMIImCl and ZnO-NPs improved oxidation signal of lorazepam about 2.17 times compared to unmodified PE. On the other hand, no specific interference for monitoring of lorazepam has been reported at surface of HMIImCl/ZnO-NPs/PE. The HMIImCl/ZnO-NPs/PE showed two-month stability for monitoring of lorazepam in aqueous solution.

References

- [1] M. Yoosefian, H. Karimi-Maleh, A. L. Sanati, A theoretical study of solvent effects on the characteristics of the intramolecular hydrogen bond in Droxidopa, *Journal of Chemical Sciences* **127**(6) (2015) 1007-1013. <https://doi.org/10.1007/s12039-015-0858-2>
- [2] R. M. Fathy, A. Y. Mahfouz, Eco-friendly graphene oxide-based magnesium oxide nanocomposite synthesis using fungal fermented by-products and gamma rays for outstanding antimicrobial, antioxidant, and anticancer activities, *Journal of Nanostructure in Chemistry* **11**(2) (2021) 301-321. <https://doi.org/10.1007/s40097-020-00369-3>
- [3] H. Bártíková, R. Podlipná, L. Skálová, Veterinary drugs in the environment and their toxicity to plants, *Chemosphere* **144**(2) (2016) 2290-2301. <https://doi.org/10.1016/j.chemosphere.2015.10.137>
- [4] A. John, L. Benny, A. R. Cherian, S. Y. Narahari, A. Varghese, G. Hegde, Electrochemical sensors using conducting polymer/noble metal nanoparticle nanocomposites for the detection of various analytes, *Journal of Nanostructure in Chemistry* **11**(1) (2021) 1-31. <https://doi.org/10.1007/s40097-020-00372-8>
- [5] P. N. Asrami, M. S. Tehrani, P. A. Azar, S. A. Mozaffari, Impedimetric glucose biosensor based on nanostructure nickel oxide transducer fabricated by reactive RF magnetron sputtering system, *Journal of Electroanalytical Chemistry* **801**(9) (2017) 258-266. <https://doi.org/10.1016/j.jelechem.2017.07.052>

- [6] B. Rezaei, A. Mokhtari, Chemiluminescence determination of promazine in human serum and drug formulations using Ru (phen)₃²⁺-Ce (IV) system and a chemometrical optimization approach, *Luminescence* **24**(3) (2009) 183-188. <https://doi.org/10.1002/bio.1093>
- [7] N. Erk, M. Kartal, Comparison of high-performance liquid chromatography and absorbance ratio methods for the determination of hydrochlorothiazide and lisinopril in pharmaceutical formulations, *Analytical Letters* **32**(6) (1999) 1131-1141. <https://doi.org/10.1080/00032719908542883>
- [8] R. Hurtubise, H. W. Latz, Fluorimetric determination of butylated hydroxy anisole in food products and packaging material, *Journal of Agricultural and Food Chemistry* **18**(3) (1970) 377-380. <https://doi.org/10.1021/jf60169a008>
- [9] C. Lacey, G. McMahon, J. Bones, M. Morrissey, J. N. Tobin, An LC-MS method for the determination of pharmaceutical compounds in wastewater treatment plant influent and effluent samples, *Talanta* **75**(4) (2008) 1089-1097. <https://doi.org/10.1016/j.talanta.2008.01.011>
- [10] S. Cheraghi, M. A. Taher, H. Karimi-Maleh, F. Karimi, M. Shabani-Nooshabadi, M. Alizadeh, A. A. Otman, N. Erk, P. V. Y. Raman, C. Karaman, Novel enzymatic graphene oxide based biosensor for the detection of glutathione in biological body fluids, *Chemosphere* **287**(1) (2022) 132187. <https://doi.org/10.1016/j.chemosphere.2021.132187>
- [11] S. Chenarani, M. Ebrahimi, V. Arabali, S. A. Beyramabadi, Determination of Lorazepam Using the Electrocatalytic Effect of NiO/SWCNTs Modified Carbon Paste Electrode as a Powerful Sensor, *Topics in Catalysis* **65**(5) (2022) 733-738. <https://doi.org/10.1007/s11244-022-01561-1>
- [12] H. Woelk, S. Schläpke, A multi-center, double-blind, randomised study of the Lavender oil preparation Silexan in comparison to Lorazepam for generalized anxiety disorder *Phytomedicine* **17**(2) (2010) 94-99. <https://doi.org/10.1016/j.phymed.2009.10.006>
- [13] M. B. Scharf, J. A. Jacoby, Lorazepam—efficacy, side effects, and rebound phenomena, *Clinical Pharmacology and Therapeutics* **31**(2) (1982) 175-179. <https://doi.org/10.1038/clpt.1982.27>
- [14] J. Ghasemi, A. Niazi, Two-and three-way chemometrics methods applied for spectrophotometric determination of lorazepam in pharmaceutical formulations and biological fluids, *Analytica Chimica Acta* **533**(2) (2005) 169-177. <https://doi.org/10.1016/j.aca.2004.11.012>
- [15] M. Ghalkhani, N. Zare, F. Karimi, C. Karaman, M. Alizadeh, Y. Vasseghian, Recent advances in Ponceau dyes monitoring as food colorant substances by electrochemical sensors and developed procedures for their removal from real samples, *Food and Chemical Toxicology* **161**(3) (2022) 112830. <https://doi.org/10.1016/j.fct.2022.112830>
- [16] P. Nasehi, M. S. Moghaddam, N. Rezaei-savadkouhi, M. Alizadeh, M. N. Yazdani, H. Agheli, Monitoring of Bisphenol A in water and soft drink products using electrochemical sensor amplified with TiO₂-SWCNTs and ionic liquid, *Journal of Food Measurement and Characterization* **16**(3) (2022) 2440-2445. <https://doi.org/10.1007/s11694-022-01321-5>
- [17] M. Alizadeh, E. Demir, N. Aydogdu, N. Zare, F. Karimi, S. M. Kandomal, H. Rokni, Y. Ghasemi, Recent advantages in electrochemical monitoring for the analysis of amaranth and carminic acid food colors, *Food and Chemical Toxicology* **163**(5) (2022) 112929. <https://doi.org/10.1016/j.fct.2022.112929>
- [18] J. A. Buledi, N. Mahar, A. Mallah, A. R. Solangi, I. M. Palabiyik, N. Qambarani, F. Karimi, Y. Vasseghian, H. Karimi-Maleh. Electrochemical quantification of mancozeb through tungsten oxide/reduced graphene oxide nanocomposite: A potential method for environmental remediation, *Food and Chemical Toxicology* **161**(3) (2022) 112843. <https://doi.org/10.1016/j.fct.2022.112843>

- [19] M. H. Karimi-Harandi, M. Shabani-Nooshabadi, R. Darabi, Simultaneous determination of citalopram and selegiline using an efficient electrochemical sensor based on ZIF-8 decorated with RGO and g-C₃N₄ in real samples, *Analytica Chimica Acta* **1203**(4) (2022) 339662. <https://doi.org/10.1016/j.aca.2022.339662>
- [20] J. Cheng, Z. Lu, X. Zhao, X. Chen, Y. Zhu, H. Chu, Electrochemical performance of porous carbons derived from needle coke with different textures for supercapacitor electrode materials, *Carbon Letters* **31**(1) (2021) 57-65. <https://doi.org/10.1007/s42823-020-00149-7>
- [21] H. Medetalibeyoğlu, An investigation on development of a molecular imprinted sensor with graphitic carbon nitride (g-C₃N₄) quantum dots for detection of acetaminophen, *Carbon Letters* **31**(6) (2021) 1237-1248. <https://doi.org/10.1007/s42823-021-00247-0>
- [22] M. Mehmandoust, N. Erk, O. Karaman, F. Karimi, M. Bijad, C. Karaman, Three-dimensional porous reduced graphene oxide decorated with carbon quantum dots and platinum nanoparticles for highly selective determination of azo dye compound tartrazine, *Food and Chemical Toxicology* **158**(12) (2021) 112698. <https://doi.org/10.1016/j.fct.2021.112698>
- [23] M. Roostaei, I. Sheikhsheh, Fabrication of a sensitive sensor for determination of xanthine in the presence of uric acid and ascorbic acid by modifying a carbon paste sensor with Fe₃O₄@ Au core-shell and an ionic liquid, *Journal of Food Measurement and Characterization* **16**(1) (2022) 731-739. <https://doi.org/10.1007/s11694-021-01200-5>
- [24] Z. Yang, Y. Zhong, X. Zhou, W. Zhang, Y. Yin, W. Fang, H. Xue, Metal-organic framework-based sensors for nitrite detection, *Journal of Food Measurement and Characterization* **16**(4) (2022), 1572-1582. <https://doi.org/10.1007/s11694-021-01270-5>
- [25] T.I. Sebokolodi, D.S. Sipuka, T.R. Tsekeli, D. Nkosi, O.A. Arotiba, An electrochemical sensor for caffeine at a carbon nanofiber modified glassy carbon electrode, *Journal of Food Measurement and Characterization* **16**(8) (2022) 2536-2544. <https://doi.org/10.1007/s11694-022-01365-7>
- [26] Y. Zhao, Y. Ma, R. Zhou, et al., Highly sensitive electrochemical detection of paraoxon ethyl in water and fruit samples based on defect-engineered graphene nanoribbons modified electrode, *Journal of Food Measurement and Characterization* **16**(8) (2022) 2596-2603. <https://doi.org/10.1007/s11694-022-01366-6>
- [27] A. A. Ensafi, H. Karimi-Maleh, S. Mallakpour, N-(3, 4-Dihydroxyphenethyl)-3, 5-dinitrobenzamide-Modified Multiwall Carbon Nanotubes Paste Electrode as a Novel Sensor for Simultaneous Determination of Penicillamine, Uric acid, and Tryptophan, *Electroanalysis* **23**(6) (2011) 1478-1487. <https://doi.org/10.1002/elan.201000741>
- [28] M. H. Karimi-Harandi, M., Shabani-Nooshabadi, R. Darabi, Cu-BTC Metal-Organic Frameworks as Catalytic Modifier for Ultrasensitive Electrochemical Determination of Methocarbamol in the Presence of Methadone, *Journal of The Electrochemical Society* **168**(9) (2021) 097507. <https://doi.org/10.1149/1945-7111/ac2468>
- [29] M. Kumar, S. S. Kirupavathy, S. Shalini, Exploration on reduced graphene oxide/strontium pyro niobate electrode material for electrochemical energy storage applications, *Carbon Letters* **31**(4) (2021) 619-633. <https://doi.org/10.1007/s42823-020-00203-4>
- [30] W. H. Daniai, N.A. Norhisham, A. F. Ahmad Noorden, Z. A. Majid, K. Matsumura, A. Iqbal, A short review on electrochemical exfoliation of graphene and graphene quantum dots, *Carbon Letters* **31**(3) (2021) 371-388. <https://doi.org/10.1007/s42823-020-00203-4>
- [31] H. Esmaeili, S. M. Mousavi, S. A. Hashemi, W. H. Chiang, S. A. Abnavi, Activated carbon@ MgO@ Fe₃O₄ as an efficient adsorbent for As (III) removal, *Carbon Letters* **31**(5) (2021) 851-862. <https://doi.org/10.1007/s42823-020-00186-2>
- [32] J. Ma, J. Yuan, W. Ming, W. He, G. Zhang, H. Zhang, Y. Cao, Z. Jiang, Non-traditional processing of carbon nanotubes, *Alexandria Engineering Journal* **61**(1) (2022) 597-617. <https://doi.org/10.1016/j.aej.2021.06.041>

- [33] R. Wu, C. Bi, X. Zhang, J. Wang, L. Wang, C. Fan, M. Wang, F. Shao, N. Li, Z. Zong, Y. Fan, Construction of two cobalt based bi-functional metal-organic frameworks for enhancing electrocatalytic water oxidation and photocatalytic disposals of hazardous aromatic dyes, *Molecular Catalysis* **505**(4) (2021) 111450. <https://doi.org/10.1016/j.mcat.2021.111450>
- [34] J. Cao, A. Li, Y. Zhang, L. Mu, X. Huang, Y. Li, T. Yang, C. Zhang, C. Zhou, Highly efficient unsupported Co-doped nano-MoS₂ catalysts for p-cresol hydrodeoxygenation, *Molecular Catalysis* **505**(4) (2021), 111507. <https://doi.org/10.1016/j.mcat.2021.111507>
- [35] G. Costa, P.A. Lopes, A. L. Sanati, A. F. Silva, M. C. Freitas, A. T. D. Almedia, M. Tavakoli, 3D Printed Stretchable Liquid Gallium Battery, *Advanced Functional Materials* **32**(27) (2022) 2113232. <https://doi.org/10.1002/adfm.202113232>
- [36] M. Al Sharabati, R. Abokwiek, A. Al-Othman, M. Tawalbeh, C. Caraman, Y. Orooji, F. Karimi, Biodegradable polymers and their nano-composites for the removal of endocrine-disrupting chemicals (EDCs) from wastewater: a review, *Environmental Research* **202**(11) (2021) 111694. <https://doi.org/10.1016/j.envres.2021.111694>
- [37] S. Akhil, A. M. M. J. Saeed, S. S. Majety, B. Mulla-muri, G. Majji, D. Bharatiya, V. S. S. Mosali, H. B. Bollikolla, B. Chandu, Cost effective biosynthetic approach for graphene exhibiting superior sonochemical dye removal capacity, *Carbon Letters* **31**(6) (2021) 1215-1225. <https://doi.org/10.1007/s42823-021-00245-2>
- [38] S. Mpelane, N. Mketo, N. Bingwa, P.N. Nomngongo, Synthesis of mesoporous iron oxide nanoparticles for adsorptive removal of levofloxacin from aqueous solutions: Kinetics, isotherms, thermodynamics and mechanism, *Alexandria Engineering Journal* **61**(11) (2022) 8457-8468. <https://doi.org/10.1016/j.aej.2022.02.014>
- [39] A. L. Sanati, A. Chambel, P. A. Lopes, T. Nikitin, R. Fausto, Laser-Assisted Rapid Fabrication of Large-Scale Graphene Oxide Transparent Conductors, *Advanced Materials Interfaces* **9**(17) (2022) 2102343. <https://doi.org/10.1002/admi.202102343>
- [40] F. Karimi, A. Ayati, B. Tanhaei, A. L. Santi, S. Afshar, A. Kardan, Z. Dabirifar, C. Karaman, Removal of metal ions using a new magnetic chitosan nano-bio-adsorbent; A powerful approach in water treatment, *Environmental Research* **203**(1) (2022) 111753. <https://doi.org/10.1016/j.envres.2021.111753>
- [41] C. Karaman, Orange peel derived-nitrogen and sulfur Co-doped carbon dots: a nano-booster for enhancing ORR electrocatalytic performance of 3D graphene networks, *Electroanalysis* **33**(5) (2021) 1356-1369. <https://doi.org/10.1002/elan.202100018>
- [42] A. Akça, O. Karaman, C. Karaman, Mechanistic insights into catalytic reduction of N₂O by CO over Cu-embedded graphene: a density functional theory perspective, *ECS Journal of Solid State Science and Technology* **10**(4) (2021) 041003. <https://doi.org/10.1149/2162-8777/abf481>
- [43] R. T. Hussain, A. S. Islam, M. Khairuddean, F. B. M. Suah, A polypyrrole/GO/ZnO nanocomposite modified pencil graphite electrode for the determination of andrographolide in aqueous samples, *Alexandria Engineering Journal* **61**(6) (2022) 4209-4218. <https://doi.org/10.1016/j.aej.2021.09.040>
- [44] H. Karimi-Maleh, F. Tahernejad-Javazmi, V. K. Gupta, H. Ahmar, M. H. Asadi, A novel biosensor for liquid phase determination of glutathione and amoxicillin in biological and pharmaceutical samples using a ZnO/CNTs nanocomposite/catechol derivative modified electrode, *Journal of Molecular Liquids* **196**(8) (2014) 258-263. <https://doi.org/10.1016/j.molliq.2014.03.049>
- [45] M. Tabrizi, S. A. Shahidi, F. Chekin, A. G. HasanSaraei, S. N. Raeisi, Reduce graphene oxide/Fe₃O₄ nanocomposite biosynthesized by sour lemon peel; using as electro-catalyst for fabrication of vanillin electrochemical sensor in food products analysis and anticancer

- activity, *Topics in Catalysis* **65**(5) (2022) 726-732. <https://doi.org/10.1007/s11244-021-01541-x>
- [46] M. Fouladgar, H. Karimi-Maleh, Ionic liquid/multiwall carbon nanotubes paste electrode for square wave voltammetric determination of methyl dopa, *Ionics* **19**(8) (2013) 1163-1170. <https://doi.org/10.1007/s11581-012-0832-7>
- [47] H. Karimi-Maleh, A. L. Sanati, V. K. Gupta, M. Yoosefian, M. Asif, A. Bahari, A voltammetric biosensor based on ionic liquid/NiO nanoparticle modified carbon paste electrode for the determination of nicotinamide adenine dinucleotide (NADH), *Sensors and Actuators B* **204**(12) (2014) 647-654. <https://doi.org/10.1016/j.snb.2014.08.037>
- [48] A.A. Ensafi, H. Karimi-Maleh, Voltammetric determination of isoproterenol using multiwall carbon nanotubes-ionic liquid paste electrode, *Drug Testing and Analysis* **3**(5) (2011) 325-330. <https://doi.org/10.1002/dta.232>
- [49] M. Alizadeh, M. Nodehi, S. Salmanpour, F. Karimi, A. L. Sanati, S. Malekmohammadi, N. Zakariae, R. Esmaeili, J. Hedayat, Properties and recent advantages of N, N'-dialkylimidazolium-ion liquids application in electrochemistry, *Current Analytical Chemistry* **18**(1) (2022) 31-52. <https://doi.org/10.2174/1573411016999201022141930>
- [50] A. Hosseini-Roudsari, S.A. Shahidi, A. Ghorbani-HasanSaraei, S. Hosseini, F. Fazeli, A new electroanalytical approach for sunset yellow monitoring in fruit juices based on a modified sensor amplified with nano-catalyst and ionic liquid, *Food and Chemical Toxicology* **168**(10) (2022) 113362 <https://doi.org/10.1016/j.fct.2022.113362>
- [51] P. Ebrahimi, S.A. Shahidi, M. Bijad, A rapid voltammetric strategy for determination of ferulic acid using electrochemical nanostructure tool in food samples, *Journal of Food Measurement and Characterization* **14**(6) (2020) 3389-3396. <https://doi.org/10.1007/s11694-020-00585-z>
- [52] A. A. Ensafi, H. Karimi-Maleh, S. Mallakpour, Simultaneous determination of ascorbic acid, acetaminophen, and tryptophan by square wave voltammetry using N-(3, 4-Dihydroxyphenethyl)-3, 5-Dinitrobenzamide-modified carbon nanotubes paste electrode, *Electroanalysis* **24**(3) (2012) 666-675. <https://doi.org/10.1002/elan.201100465>
- [53] H. Karimi-Maleh, F. Tahernejad-Javazmi, M. Daryanavard, Electrocatalytic and simultaneous determination of ascorbic acid, nicotinamide adenine dinucleotide and folic acid at ruthenium(II) complex-ZnO/CNTs nanocomposite modified carbon paste electrode, *Electroanalysis* **26**(5) (2014) 692-970. <https://doi.org/10.1002/elan.201400013>
- [54] J. B. Raoof, R. Ojani, H. Karimi-Maleh, M. R. Hajmohammadi, P. Biparva, Multi-wall carbon nanotubes as a sensor and ferrocene dicarboxylic acid as a mediator for voltammetric determination of glutathione in hemolysed erythrocyte, *Analytical Methods* **3**(11) (2011) 2637-2643. <https://doi.org/10.1039/C1AY05031A>
- [55] R. Darabi, M. Shabani-Nooshabadi, Development of an amplified nanostructured electrochemical sensor for the detection of cefixime in pharmaceuticals and biological samples, *Journal of Pharmaceutical and Biomedical Analysis* **212**(5) (2022) 114657. <https://doi.org/10.1016/j.jpba.2022.114657>